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Field Dose-Response Approach (DRAC-"Dose-Resposta para Avaliação da Contaminação") as a Tool for Environmental Mercury Contamination Assessment in Fish

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Methylmercury (MeHg) is a well known human neurotoxin (Clarkson, 1994). The general population is primarily exposed to MeHg through fish consumption (WHO, 1990; Clarkson, 1994; U.S. EPA, 2001). It has been demonstrated that Hg usually accumulates in fish tissues as MeHg, from inorganic Hg sources (Huckabee et al., 1979). Traditionally, the United States Environmental Protection Agency (U.S. EPA) has expressed, in its section 304(a), the water quality criteria guidance, to protect human health, as the pollutant concentration in ambient surface water. In January 2001, the U.S. EPA (2001) concluded that it is more appropriate, at this time, to derive a fish tissue (including shellfish) residue water quality criterion for MeHg rather than a water column-based water quality criterion. This criterion is based directly on the dominant human exposure route to MeHg.

Many factors have been considered as important in the bioaccumulation and/or biomagnification of Hg in fish. Among them, the Hg load-dependent factors in the aquatic environment, specially those related to Hg in sediments and environmental conditions, like bio-production (Häkanson, 1980; 1991); as well as local biota's physiological-dependent factors, like size, length, age and metabolic rate (Phillips, 1980; WHO, 1990); and, in addition, the food-chain characteristics (Cabana et al, 1994). In general, the Hg levels in fish muscles show large inter-individual variabilities, resulting in very high values for relative standard deviation (U.S. EPA, 1999). Hg levels in fish for spatial and/or temporal comparisons have been normalized by the average Hg content in 1 Kg of fish (as pike) (Johnels et al., 1967; Häkanson, 1991), by using only fish of one year old (Post et al, 1996), or a specific length (Scruton, et al., 1994), or a specific weight (Watras et al., 1998). Castilhos et al (2001) suggested a field dose-response approach as a tool for assessing the environmental Hg contamination, through the mercury analysis in fish. Doseresponse approach is usually used for analysis of data from laboratory or epidemiologic studies. However, we have proposed its use for assessment of Hg contamination in fish sampled from field. This methodology was used to assess the Hg contamination, in the Tapajós River Region, caused by gold mining activities. The results showed different daily uptake doses by Tucunaré (*Cichla* spp.) between non-contaminated and contaminated areas (~4.0 times), and such differences could be attributed to different Hg loading rates between the studied areas.

We have tested the applicability of the proposed methodology (DRAC) by using the literature data. Seven species of fishes from 8 different aquatic ecosystems, including lentic and lotic freshwater, estuarine and marine ecosystems, that were chosen from scientific publications, in which Hg levels in muscles and estimated age and/or measured length for individual specimens, were available. The objectives of this work were: (i) to establish and compare the dose-response relationship for Hg accumulation by different fish species, from several ecosystem and collection time (ii) to estimate and compare the daily Hg uptake rate by those different fishes; (iii) to estimate and compare the potential exposure time necessary for Hg accumulation to reach 500µg.Kg⁻¹, the limit concentration for human consumption, adopted in many countries, and, (iv) to present this approach, in addition to that expressed by U.S. EPA, for water quality criteria for methyl-mercury.

MATERIALS AND METHODS

Seven species of fish, from 8 different aquatic ecosystems, including lentic and lotic freshwater, estuarine and marine ecosystems were chosen from scientific publications, in which Hg levels in muscles and estimated age or measured length for individual specimens were available. For the literature that shows the relationship between the Hg levels in tissues and the size and/or age of fish, in a graphical form, the program "GIF Coordinates" was used for the coordinate acquisition (Schneider, 2000).

The "DRAC" methodology was described in a previous work (Castilhos et al., 2001), and a brief description is presented as follows. The dose-response relationship has the competence to absorb inter-individual variabilities. The responses are of two kinds: Quantal and quantitative. In a quantitative test, an organism either or not shows the response under study. Thus, a certain percentage of test organisms will show the response within some stated conditions. In a quantitative or graded test, each organism responds to a distinct degree. The Quantal test are designed to estimate the concentration of a test material that affects 50% of the test organisms, the mean effective dose (ED 50% or ED50). One must choose the effect to be observed. Thus, this is a quantal rather than a graded response, since the specific effect is either present or absent. The ED50, for accumulation of Hg by fish, indicate the exposure time necessary to attain those tissue concentration levels by half of the exposed individuals. Some methods are used to calculate ED50, and, among them, there is the "probit" method (American Public Health Association, 1985; Ross and Gilman, 1985). The potential exposure times were inferred from estimated age and were transformed in their logarithms. The frequency of responses were transformed in "probit" units. The D_{50} for accumulation of Hg by fish (accumulation dose 50 or AD_{50}) indicates the exposure time necessary to attain those tissue concentration levels by half of the exposed individuals. This resulting time can be related to response, as

follows: t exposure * C = constant (adapted from Dämgen & Grünhage, 1998); in which a certain response (K, constant) can be achieved from a exposure time (t exposure) and the concentration in the aquatic environment C; such concentration will result as a potential dose or daily uptake rate (DUR), expressed in $\mu g.Kg^{-1}.d^{-1}$. From these results one could estimate the exposition time to reach either 300 $\mu g.Kg^{-1}$ or 500 $\mu g.Kg^{-1}$ and compare the contamination magnitude (or bioavailability) among different aquatic ecosystems.

RESULTS AND DISCUSSION

In environmental hazard analysis, the ecological effect of a specific substance on an organism or group of organisms in an aquatic system can be described as a function of the dose during a defined period of time. The direct bioaccumulation factor (BAF) or bioconcentration factor (BCF) of Hg is defined as the ratio of Hg concentration in fish tissue to the Hg concentration in water. The indirect bioaccumulation or biomagnification is the accumulation of a chemical in a given species according to its trophic levels in the food chain (Bruggeman, 1982). Bioaccumulation and/or biomagnification are the most direct study of mercury reaction at the environmental interface of the aquatic organisms. In this particular context, the ecological effect is defined as Hg-content in fish.

Although the bioaccumulation/biomagnification processes have not been traditionally interpreted as a pharmacological/toxicological "effect", one could suggest that as higher the internal dose (bioaccumulation), the higher is the potential aquatic risk. For instances, MeHg has been classified as clastogen substance and toxic effects such as icthyomutagenesis have been associated to MeHg exposure and dose intensity (probabilistic risk) (Schoeny, 1997). Moreover, several effects show a positive correlation with exposure but a causal-effect relationship is a difficult task to access, but bioaccumulation/biomagnification processes, which have to have, at least, exposure conditions for a determinant chemical agent.

The U.S. EPA (2001) concluded that it is more appropriate to derive a fish tissue (including shellfish) residue water quality criterion for MeHg rather than a water column-based water quality criterion, by using the following equation.

$$TRC = \underline{BW \times (RfD - RSC)}_{\Sigma^4_{i=2} FI_i}$$

Where:

TRC = Fish tissue residue criterion (mg methyl-mercury/ kg fish) for freshwater and estuarine fish

RfD = Reference dose (based on non-cancer human health effects) of 0.0001mg/kg body weight-day

RSC = Relative source contribution (subtracted from RfD to account for marine fish consumption) estimated to be $2.7x10^{\text{-}5}$ mg methyl-mercury / kg body weight-day

 $(RfD - RSC) = (1x \ 10^{-4} - 0.27x10^{-4}) = 0.73x10^{-4} \text{ mg methyl-mercury / kg}$ body weight-day or 0.073 µg methyl-mercury / kg body weight-day

BW = Human body weight (default value of 70 kg) (for adults)

FI = Fish intake at trophic level (TL) i (i = 2, 3, 4); total intake is 0.0175 kg fish / day for general adult population.

The resulting Fish Residue Criterion is 0.3 mg methylmercury/kg fish. This is the concentration in fish tissue that should not be exceed based on a total fish and shellfish consumption-weighted rate of 0.0175 kg fish.d⁻¹.

The Reference Dose (RfD) is an estimate (with total uncertainties spanning up to five orders of magnitude) of a daily exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious health effects during a lifetime. In the risk assessment discussion U.S. EPA uses the National Research Council (NRC)-recommended Benchmark Dose Lower Limit (BMDL) of 58 ppb mercury in cord blood as an example of RfD calculation. By using an onecompartment model, the methylmercury ingestion dose that corresponds to a cord blood level of 58ppb, is 1.081 µg.Kg⁻¹ bw.d⁻¹. Considering the uncertainties factors, the RfD for methylmercury resulted in 0.1 µg.kg⁻¹ bw.d⁻¹ or 1x10⁻⁴ mg.kg⁻¹ bw.d⁻¹. Based on available data, human exposures to methylmercury from all media sources except freshwater/estuarine and marine fish are negligible, both in comparison with exposures from fish and compared with RfD. However, ingestion of marine fish is a significant contributor to the total methylmercury exposure. For the methylmercury criterion, the RSC is the estimated exposure from marine fish intake. This is substracted from the RfD when calculating the water quality criterion, and represents almost 30% of the RfD.

It has been repeatedly shown that mercury in fish accumulates throughout the lifetime of the individual, and BAF values for given species vary as a function of ages of the animals examined. Most of the water body-specific BAFs and resulting level distributions, are based on "opportunity" (whatever you catch, you include), composite samples and do not report age or size of individuals sampled (U.S EPA, 2001). Then, one only can sample very small fish specimens to show adequacy to the TRC value.

The results are presented in Table 1, which displays the popular name of the fish, the local and time of sampling, food habit, number of collected specimens, observed effect (OE), daily uptake rate estimate (DUR), estimate of time of exposition to reach 500 µg.Kg⁻¹, and references. Comparing the OE values (mean Hg concentration in µg.kg⁻¹) in the population sampled, for piscivorous and zooplanktivorous fishes, one could suggest the following crescent order of contamination or crescent contamination magnitude/bioavailability: Sepetiba Bay (corvina) < Ilha Grande Bay (corvina) = Guanabara Bay (corvina) = lagoons-Santarém (tucunaré) < Conceição Lagoon (corvina) < Tongue River-reservoir (white crapie) < Tongue River-reservoir (walleye) = Tongue River-reservoir (pike) = Tapajós River (tucunaré) < Tongue

River- reservior (sauger) < Gulf of Trieste (conger-conger) < Tucuruí River-reservoir (tucunaré). However, considering the daily uptake rate estimates and/or the time necessary for half of the specimens to reach 500µg.Kg⁻¹, the order of contamination is altered to: Tongue River-reservoir (white crapie) < Conceição Lagoon (corvina) < Ilha Grande Bay < Guanabara Bay < Sepetiba Bay < Tongue River-reservoir (pike = sauguer= walleye) = lagoons-Santarém (tucunaré) < Tapajós River (tucunaré) < Gulf of Trieste (conger-conger) < Tucuruí River-reservoir (tucunaré).

Considering TRC, sauger (Tongue River), tucunaré (Tucuruí Reservoir) and congerconger (Gulf of Trieste) showed Hg levels above the limit. However, looking at the estimated daily uptake rates, northern pike, sauger and walleye (Tongue River) and tucunaré (lagoons=Santarém) showed the same value: $0.2\mu g.kg^{-1}d^{-1}$. The estimated daily uptake rates for conger-conger (Gulf of Trieste), tucunaré from Tapajós River and tucunaré from Tucuruí Reservoir resulted in 2 times, 4 times and 8 times higher than $0.2\mu g.kg^{-1}d^{-1}$, respectively.

Considering data from the work relative to *Micropogonias furnieri*, when the average of Hg levels are normalized with respect to the average weight of the fish (not presented here), no alteration in the order of contamination is achieved, as compared to the order obtained by using DRAC. On the other hand, the zooplanktivorous fish from Tongue River reservoir became, as expected, the less contaminated specie compared to carnivorous and piscivorous fish. For the Tucuruí system the possible differences in the tucunaré growth rate should be investigated, since the literature data indicate the existence of dwarf species in reservoirs (Doyon et al., 1998).

Not surprising, some data may be lost during their transference from the plots of original articles to the "GIF coordinates", and the results might be improved by using the original data. However, the present objective was to test the applicability of the dose-response approach as a tool for environmental assessment mercury contamination by using the literature data. One of the ways to try to establish the weakest, most sensitive and uncertain part of a model is to conduct sensitivity tests. The selection of dose-response models (in this work, probit) and the effect of uncertainties in these models should be evaluated. Indeed, the results might not represent, actually, the magnitude of mercury contamination/availability among sites, because there are significant differences in temporal sampling as well as in analytical procedures.

We suggest that, in addition to TRC (0.3 mg methylmercury / kg fish) for specific fish specie, one could estimate the daily uptake rate, which may express the bioavailability of mercury in a defined aquatic ecosystem, and compare the time necessary to attain the TRC value. For using the dose-response approach, one should collect about 30 individuals (within intervals related with percentage of the specie-specific maximum length resulting three points in the dose-response relationship). Such approach might permit an integrate comparison among different aquatic ecosystems using the same or different fish species. The DRAC is a simple and fast

estimated daily uptake rate (DUR; µg.Kg⁻¹.d⁻¹), estimated time of exposure to attain either 300µg.Kg⁻¹ (T300, years) or 500µg.Kg⁻¹ Table 1. Popular and scientific names, food habit, locality, date and number of fish collected (N), observed effect (OE;µg.Kg⁻¹) (T500, years) and references.

Esox lucius P Tongue River- Reservoir (EUA)' 1978 56 300 Stizostedion P Tongue River- Reservoir (EUA)' 1978 56 300 Stizostedion vitreum P Tongue River- Reservoir (EUA)' 1978 26 300 Stizostedion vitreum P Tongue River- Reservoir (EUA)' 1978 26 300 Micropogonias C Guanabara Bay (RJ, BR)² 1992 56 100 furnieri Micropogonias C Ilha Grande Bay (RJ, BR)² 1990-1991 57 100 furnieri Micropogonias C Conceição Lagoon (SC, BR)² 1990-1991 42 130 furnieri Cichla spp P LagoonsSantarém (PA, BR)³ 1992 28 100 Cichla spp P Tucurui River- Reservoir (PA, BR)³ 1995 61 1000 Cichla spp P Tucurui River- Reservoir (PA, BR)³ 1995 61 1000 Cichla spp P Gulf of Trieste (Slovenia)² 1995-1996 25 610	Donnler name	Coientific name	*U2	Toooliter	Doto	7	OE	DUR	T300	T500
Esox lucius P Tongue River- Reservoir (EUA)' 1978 56 Stizostedion P Tongue River- Reservoir (EUA)' 1978 31 canadense Stizostedion vitreum P Tongue River- Reservoir (EUA)' 1978 26 Pomoxis annularis Z Tongue River- Reservoir (EUA)' 1978 36 Micropogonias C Guanabara Bay (RJ, BR)² 1992 56 furnieri Micropogonias C Ilha Grande Bay (RJ, BR)² 1990-1991 57 furnieri Micropogonias C Conceição Lagoon (SC, BR)² 1990-1991 42 furnieri Cichla spp P Lagoons – Santarém (PA, BR)³ 1992 28 Cichla spp P Tucuruí River- Reservoir (PA, BR)³ 1995 61 Cichla spp P Tucuruí River- Reservoir (PA, BR)³ 1995 61 Cichla spp P Tucuruí River- Reservoir (PA, BR)³ 1995 61	1 Opulai manno		111	Locality	Date	ζ,	$(\mu g.Kg^{-1})$	$(\mu g. Kg^{-1}. d^{-1})$	(years)	(years)
Stizostedion P Tongue River- Reservoir (EUA) ⁷ 1978 31 canadense Stizostedion vitreum P Tongue River- Reservoir (EUA) ⁷ 1978 26 Pomoxis annularis Z Tongue River- Reservoir (EUA) ⁷ 1978 36 Micropogonias C Guanabara Bay (RJ, BR) ² 1990-1991 57 furnieri Micropogonias C Ilha Grande Bay (RJ, BR) ² 1990-1991 57 furnieri Micropogonias C Conceição Lagoon (SC, BR) ² 1990-1991 60 furnieri Cichla spp P Lagoons – Santarém (PA, BR) ³ 1992 28 Cichla spp P Tucuruí River- Reservoir (PA, BR) ³ 1995 61 Conser conser Conser Conser	Northern Pike	Esox lucius	Ъ	Tongue River- Reservoir (EUA) ¹	1978	99	300	0.2	4.1	6.8 5
Canadense Stizostedion vitreum P Tongue River-Reservoir (EUA) ⁷ 1978 26 Pomoxis annularis Z Tongue River-Reservoir (EUA) ⁷ 1978 36 Micropogonias C Guanabara Bay (RJ, BR) ² 1992 56 furnieri Micropogonias C Ilha Grande Bay (RJ, BR) ² 1990-1991 57 furnieri Micropogonias C Sepetiba Bay (RJ, BR) ² 1990-1991 60 furnieri Micropogonias C Conceição Lagoon (SC, BR) ² 1990-1991 42 furnieri Cichla spp P Lagoons –Santarém (PA, BR) ³ 1992 28 Cichla spp P Tucuruí River-Reservoir (PA, BR) ³ 1995 61 Conser conser Conser Conser	Sauger	Stizostedion	Д	Tongue River- Reservoir (EUA) ¹	1978	31	350	0.2	4.1	6.8 5
Stizostedion vitreum P Tongue River-Reservoir (EUA)' 1978 26 Pomoxis annularis Z Tongue River-Reservoir (EUA)' 1978 36 Micropogonias C Guanabara Bay (RJ, BR)² 1992 56 furnieri Micropogonias C Ilha Grande Bay (RJ, BR)² 1990-1991 57 furnieri Micropogonias C Sepetiba Bay (RJ, BR)² 1990-1991 60 furnieri Micropogonias C Conceição Lagoon (SC, BR)² 1990-1991 42 furnieri Cichla spp P Lagoons –Santarém (PA, BR)³ 1992 28 Cichla spp P Tucuruí River-Reservoir (PA, BR)³ 1995 61 Conser conser P Culf of Trieste (Slovenia)² 1995-1996 25		canadense								
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Micropogonias C Guanabara Bay (RJ, BR)² 1992 56 furnieri Micropogonias C Ilha Grande Bay (RJ, BR)² 1990-1991 57 furnieri Kicropogonias C Sepetiba Bay (RJ, BR)² 1990-1991 60 furnieri Micropogonias C Conceição Lagoon (SC, BR)² 1990-1991 42 furnieri P Lagoons – Santarém (PA, BR)³ 1992 28 Cichla spp P Tagajós River (PA, BR)³ 1992 41 Cichla spp P Tucuruí River- Reservoir (PA, BR)³ 1995 61 Conser conser P Gulf of Trieste (Slovenia)² 1995-1996 25	White crapie	Pomoxis annularis	Z	Tongue River- Reservoir (EUA) ¹	1978	36	200	0.05	16.4	27.0 5
Micropogonias C Ilha Grande Bay (RJ, BR)² 1990-1991 57 furnieri Sepetiba Bay (RJ, BR)² 1990-1991 60 furnieri C Conceição Lagoon (SC, BR)² 1990-1991 42 furnieri P Lagoons – Santarém (PA, BR)³ 1992 28 Cichla spp P Tapajós River (PA, BR)³ 1992 41 Cichla spp P Tucuruí River- Reservoir (PA, BR)³ 1995 61 Conser conser P Gulf of Trieste (Slovenia)² 1995-1996 25	Corvina	Micropogonias furnieri	C	Guanabara Bay (RJ, BR) 2	1992	26	100	0.15	5.5	9.1 6
MicropogoniasCSepetiba Bay (RJ, BR)²1990-199160furnieriMicropogoniasCConceição Lagoon (SC, BR)²1990-199142furnieriPLagoons –Santarém (PA, BR)³199228Cichla sppPTapajós River (PA, BR)³199241Cichla sppPTucuruí River- Reservoir (PA, BP)³199561Conper conserPGulf of Trieste (Slovenia)²1995-199625	Corvina	Micropogonias furnieri	C	Ilha Grande Bay (RJ, BR)²	1990-1991	57	100	0.13	6.3	10.5 6
Micropogonias C Conceição Lagoon (SC, BR) ² 1990-1991 42 furnieri Cichla spp P Lagoons –Santarém (PA, BR) ³ 1992 28 Cichla spp P Tapajós River (PA, BR) ³ 1992 41 Cichla spp P Tucurui River- Reservoir (PA, 1995 61 BR) ⁴ 1995 61	Corvina	Micropogonias furnieri	C	Sepetiba Bay (RJ, BR) ²	1990-1991	09	80	0.10	8.2	13.7 6
Cichla spp P Lagoons –Santarém (PA, BR) ³ 1992 28 Cichla spp P Tapajós River (PA, BR) ³ 1992 41 Cichla spp P Tucurui River- Reservoir (PA, 1995 61 BR) ⁴ Conser conser P Gulf of Trieste (Slovenia) ⁷ 1995-1996 25	Corvina	Micropogonias furnieri	C	Conceição Lagoon (SC, BR)²	1990-1991	42	130	0.08	10.3	17.0^{6}
Cichla spp P Tapajós River (PA, BR) ³ 1992 41 Cichla spp P Tucurú River- Reservoir (PA, 1995 61 BR) ⁴ Conser conser P Gulf of Trieste (Slovenia) ⁷ 1995-1996 25	Tucunaré	Cichla spp	Ь	Lagoons –Santarém (PA, BR) ³	1992	28	100	0.2	4.1	8.9
Cichla spp P Tucuruí River- Reservoir (PA, 1995 61 BR) ⁴ Conser conser P Gulf of Trieste (Slovenia) ⁷ 1995-1996 25	Tucunaré	Cichla spp	Ч	Tapajós River (PA, BR) ³	1992	41	300	8.0	1.0	2.0
P Gulf of Trieste (Slovenia) 7 1995-1996 25	Tucunaré	Cichla spp	Ы	Tucuruí River- Reservoir (PA, BR) ⁴	1995	61	1000	1.65	0.5	0.8 5
CT OCCI (DUC) COURT TO THE COUR	TANK PORTON PER	Conger conger	Ь	Gulf of Trieste (Slovenia) 7	1995-1996	25	610	0.4	2.1	3.4

* P = piscivorous; C= carnivorous benthic; Z = zooplanktivorous

Phillips et al., 1980; ²Kherig, 1992 (data non-normalized); ³Castilhos et al., 2001; ⁴Porvari, 1995; ⁵Castilhos & Lima, 2001; ⁶Castilhos, 1999; 7-Horvat et al.,1999.

methodology and can be applied to any data bank including spatial and temporal contamination assessment for environmental contaminants that show bioaccumulation.

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REFERENCES

- American Public Health Association (1985) Part 800 Toxicity test methods for aquatic organisms, In: 16th Edition, Greenberg AE, Trussell RE, Clesceri, L. and Frason MAH (ed) Standard Methods for Examination of Water and Wastewater. Baltimore, Maryland, USA, p. 689-819.
- Bruggeman WA (1982) Hidrophobic interactions in the aquatic environment. In: Hutzinger O (ed) The Handbook of environmental chemistry, vol 2. Spring-Verlag, Germany, p 205.
- Cabana G, Tremblay A, Kaff J, Rasmussen JB (1994) Pelagic food chain Structure in Ontario Lakes: A determinant of mercury levels in lake trout (Salvelinus namaycush). Canadian J Fish Aquat Sci 51:381-389.
- Castilhos ZC (1999) Estimativa da taxa de captação diária de Hg por Micropogonias furnieri a partir da interrelação dose-resposta em quatro estuários brasileiros. "Workshop" Efeitos de Poluentes em Organismos Marinhos, UFF/FAPERJ; 08 a 10 de novembro de 1999, p.22.
- Castilhos ZC & Bidone ED (2000) Mercury biomagnification in the icthyofauna of the Tapajós River Region, Brazil. Bull Environ Contam Toxicol 64:5, 693-700.
- Castilhos ZC, Bidone ED, Hartz, S (2001) Bioaccumulation of Hg by Tucunaré (Cichla ocellaris.) from Tapajós River Region, Brazilian Amazon. A Field Dose-Response Approach. Bull Environ Contam Toxicol 66:631-637.
- Castilhos ZC and Lima C (2001) Interrelação dose-resposta para a avaliação da magnitude da contaminação mercurial em peixes. VI Congresso de Geoquímica dos Países de Língua Portuguesa- Faro, Portugal, 9 a 12 de abril de 2001.
- Clarkson TW (1994) The toxicology of mercury and its compounds. In: Watras CJ, Huckabee JW (ed) Mercury pollution: Integration and synthesis. Lewis Publishers, Boca Raton, Florida, USA, p. 631-640.
- Dämgen U and Grünhage L (1998) Response of a grassland ecosystem to air pollutans. V. A toxicological model for the assessment of dose-response relationship for air pollutants and ecosystems. Environ Pollut 101:375-380.
- Driscoll CT, Schofield CL, Munson R K, YAN C, KOLSAPPLE JG (1994)The mercury cycle and fish in the Adirondack lakes. Environ Sci Technol 28:136-143.
- Doyon J-F; Bernatchez L, Gendron M, Verdon R, Fortin R (1998) Comparison of normal and dwarf populations of lake whitefish (*Coregonus clupeaformis*) with reference to hydroelectric reservoirs in northern Quebec. Arch Hydrobiol Spec Issues Advanc Limnol 50:97-108.

- Horvat, M et al (1999) Mercury in contaminated coastal environments; a case study: the Gulf of Trieste. Sci Tot Environ 237/238:43-56.
- Huckabee JW, Elwood JW, Hildebrand SG (1979) Accumulation of mercury in freshwater biota. In: Nriagu JO (ed). The biogeochemistry of mercury in the environment, Elsevier, North Holland, Amsterdam, p. 277-302.
- Håkanson, L (1980) An ecological risk index aquatic pollution control. A sedimentological approach. Water Res:975-1001.
- Håkanson, L (1991) Mercury in fish-geographical and temporal perspectives. Wat Air Soil Pollut 55:159-177.
- Johnels AGT, Westemark T, Berg W, Persson PI and Sjonstrand (1967) Pike (Esox lucius L) and some other aquatic organisms in Sweden as indicators of mercury contamination in the environment. Oikos 18:232-33.
- Kherig, H. (1992) Estudo comparativo dos níveis de concentração de mercúrio total em corvinas (Micropogonias furnieri) de quatro estuários brasileiros. Dissertação de mestrado Rio de Janeiro PUC.
- Phillips DJH (1980) The effects of age (size, weight) on trace metals in aquatic biota. In: Mellanby K (ed) Quantitative aquatic biological indicators. Applied Science Publishers, Essex, England.
- Phillips GR, Lenhart TE, Gregory RW (1980) Relation between trophic position and mercury accumulation among fishes from the Tongue river reservior, Montana. Environ Res 22:73-80.
- Porvari P (1995) Mercury levels of fish in Tucuruí hydroeletric reservior and in river Mojú in Amazonia, in the state of Pará, Brazil. Sci Total Environ 175:109-117.
- Post JR, Vandenbos R and Mcqueen J (1996) Uptake rates of food-chain and waterborne mercury by fish: field measurements, a mechanistic model, and an assessment of uncertainties. Canadian J Fish Aquat Sci 53:395-407.
- Ross EM & Gilman AG (1985) Pharmacodynamics: Mechanisms of drug action and the relationship between drug concentration and effec. In: Goodman A, Gilman A, Goodman LS, Rall TW, Murad F (ed) 17th Goodman and Gilman's The Pharmacological Basis of Therapeutics MacMillan Publishing Company, New York, NY, p.35-48.
- Schneider, C (2000) GIF Coordinates software developed at The University of Utah. Schoeny, R (1996) Use of genetic toxicology data in U S EPA risk assessment: The mercury study report as an example. Environ Health Perspect 104:663-673.
- Scruton DA, Petticrew EL, Ledrew LJ, Anderson MR, Williams UP, Bennet BA, Hill, EL (1994) Methylmercury levels in fish tissue from three reservoir systems in Insular Newfoundland, Canadá. In: Watras CJ, Huckabee JW Mercury Pollution: Integration and Synthesis. Lewis Publishers, USA, 727p.
- U.S. EPA (1999) The national survey of mercury concentrations in fish data base summary 1990-1995, Washington DC, 210p.
- U.S. EPA (2001) Water Quality Criteria- Methylmercury. www.epa.gov.
- Watras CJ, Back RC, Halvorsen S, Hudson RJM, Morrison KA, and Wente SP (1998) "Bioaccumulation of mercury in pelagic freshwater food webs". Sci Tot Environ 19:183-208.
- WHO (1990) Environmental Health Criteria 101: Methylmercury. Geneva, World Health Organization.